

Two New Disulfated Triterpenoids from *Zygophyllum fabago*

by Saleha S. Khan^{a)}, Afsar Khan^{b)}, Amir Ahmed^{a) d)}, Viqar U. Ahmad^{*a)}, Umar Farooq^{b)},
Saima Arshad^{a)}, Sadia Bader^{a)}, Aqib Zahoor^{a)}, Imran N. Siddiqui^{a)}, Bilge Sener^{c)}, and
Nurgun Erdemoglu^{c)}

^{a)} H.E.J. Research Institute of Chemistry, International Center for Chemical and Biological Sciences,
University of Karachi, Karachi-75270, Pakistan (phone: +92-21-34819020; fax: +92-21-34819018-9;
e-mail: vuahmad@hotmail.com)

^{b)} Department of Chemistry, COMSATS Institute of Information Technology, Abbottabad-22060,
Pakistan

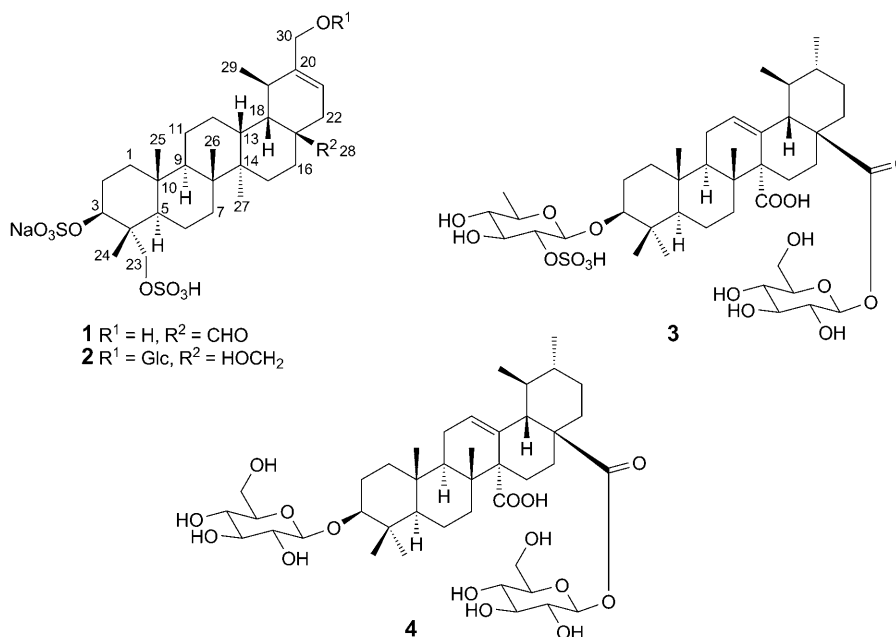
^{c)} Department of Pharmacognosy, Faculty of Pharmacy, Gazi University, Ankara-06330, Turkey

^{d)} Pharmaceutical Division, Pakistan Council of Scientific and Industrial Research Laboratories
Complex, Karachi 75280, Pakistan

From the aerial parts of *Zygophyllum fabago*, two new monosodium salts of sulfated derivatives of ursolic acid, along with two known quinovic acid glycosides were isolated. The structures of the new compounds were determined as (3 β ,4 α)-3,23,30-trihydroxyurs-20-en-28-al 3,23-di(sulfate) sodium salt (1:1) (**1**) and of (3 β ,4 α)-3,23,28-trihydroxyurs-20-en-30-yl β -D-glucopyranoside 3,23-di(sulfate) sodium salt (1:1) (**2**) with the molecular formula C₃₀H₄₇NaO₁₀S₂ and C₃₆H₅₉NaO₁₅S₂, respectively. The structures of the known compounds were 3-O-(2-O-sulfo- β -D-quinovopyranosyl)quinovic acid 28- β -D-glucopyranosyl ester (**3**) and 3-O-(β -D-glucopyranosyl)quinovic acid 28- β -D-glucopyranosyl ester (**4**) (quinovic acid = (3 β)-3-hydroxyurs-12-ene-27,28-dioic acid). The structures of all these compounds were determined by using 1D- and 2D-NMR spectroscopic techniques.

Introduction. – Chemical studies carried out on the family Zygophyllaceae have revealed the occurrence of important secondary metabolites such as sulfated triterpenoid saponins [1]. In an ongoing search for the bioactive compounds from Zygophyllaceae plants, the EtOH extract of *Zygophyllum fabago* was selected for investigation. From previous studies, several unusual disulfated ursane derivatives with a double bond at C(20)=C(21) have been reported which might have good potential in various biological activities. Compounds isolated in the present study include the two new disulfated triterpenoids **1** and **2**, as well as two known compounds 3-O-(2-O-sulfo- β -D-quinovopyranosyl)quinovic acid 28- β -D-glucopyranosyl ester (= zygophyloside E; **3**) [2] and 3-O-(β -D-glucopyranosyl)quinovic acid 28- β -D-glucopyranosyl ester (= guettarda saponin II; **4**) [3] (quinovic acid = (3 β)-3-hydroxyurs-12-ene-27,28-dioic acid).

Results and Discussion. – Compound **1** was isolated as a white amorphous powder. Its molecular formula was determined as C₃₀H₄₇NaO₁₀S₂ on the basis of HR-FAB-MS which showed the quasi-molecular-ion peak at m/z 653.2435 ($[M - H]^-$). The IR spectra of **1** showed absorption bands for OH groups (3408 cm⁻¹), C–H stretch (2925 cm⁻¹), an aldehyde C=O (1710 cm⁻¹), an olefinic bond (1627 cm⁻¹), and a band

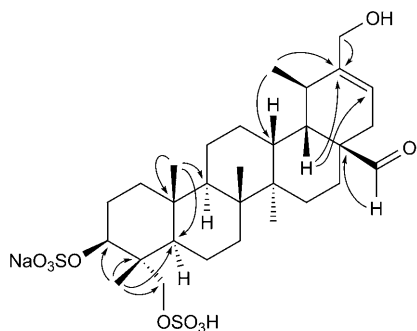


typical for a sulfate stretch (1122 cm^{-1}). Acid hydrolysis of **1** followed by treatment with BaCl_2 yielded barium sulfate, hence established the presence of a sulfate residue in **1** [4]. The $^1\text{H-NMR}$ spectrum (*Table*) exhibited an olefinic H-atom resonance at $\delta(\text{H})$ 5.56 (*d*, $J = 6.5\text{ Hz}$, H–C(21)), two CH groups at $\delta(\text{H})$ 4.38 (*dd*, $J = 4.5, 11.5\text{ Hz}$) and 9.42 (*s*), two CH₂ groups at $\delta(\text{H})$ 3.97 (*m*, H_a–C(23) overlapped with H_a–C(30)), 3.77 (*d*, $J = 9.5\text{ Hz}$, H_b–C(23)), and $\delta(\text{H})$ 3.94 (*d*, $J = 12.5\text{ Hz}$, H_b–C(30)), four tertiary Me groups at $\delta(\text{H})$ 0.76 (*s*, Me(24)), 0.93 (*s*, Me(25)), 0.94 (*s*, Me(26)), and 1.02 (*s*, Me(27)), and one secondary Me group at $\delta(\text{H})$ 1.08 (*d*, $J = 6.5\text{ Hz}$, Me(29)). In addition, the $^{13}\text{C-NMR}$ spectra (*Table*) displayed one oxygenated CH group signal at $\delta(\text{C})$ 80.8, two oxygenated CH₂ groups at $\delta(\text{C})$ 64.8 and 70.0, and five Me groups at $\delta(\text{C})$ 13.3, 14.8, 16.5, 17.2, and 23.8. The chemical shift of the CH(18) group, usually found at $\delta(\text{C})$ 50.7–51.1 in ursane-type triterpenoids [5], was shifted upfield to $\delta(\text{C})$ 41.3 in compound **1** and unambiguously assigned by a HMBC to H–C(19). Accordingly, **1** was suggested to contain an urs-20-ene skeleton and a disulfate group similar to zygofaboside A [6], except for the presence of an additional CH₂O group and the absence of a glycosidic ester linkage. The additional CH₂O group was placed at C(30) of **1** as a consequence of the low-field-shifted signals of the olefinic bond at $\delta(\text{C})$ 148.1 (C(20)) and 118.3 (C(21)), and the HMBC (*Fig. 1*) H–C(21) ($\delta(\text{H})$ 5.56)/C(30) ($\delta(\text{C})$ 64.8). The $^1\text{H-NMR}$ signal at $\delta(\text{H})$ 9.42 was consistent with the presence of an aldehyde H-atom in conjunction with the $^{13}\text{C-NMR}$ signal at $\delta(\text{C})$ 209.0. This spectroscopic evidence suggested that a CHO group could be best accommodated at the C(28) position. On the basis of the above evidences, the structure of **1** was assigned as ($3\beta,4\beta$)-3,23,30-trihydroxyurs-20-en-28-al 3,23-di(sulfate) sodium salt (1 : 1), which is a new natural compound.

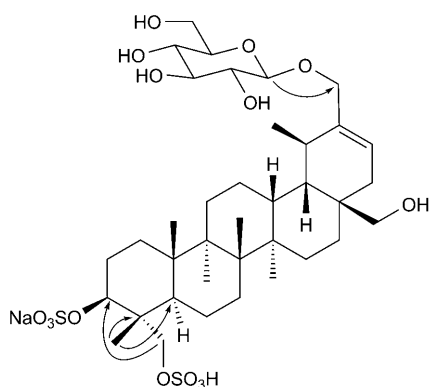
Table. ^{13}C - and ^1H -NMR Data (125 and 500 MHz, resp., CD_3OD) of **1** and **2**. δ in ppm, J in Hz.

	1		2	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
$\text{CH}_2(1)$	1.75 (br. s), 0.98 (br. s)	39.4	1.76 (br. s), 0.99 (br. s)	39.4
$\text{CH}_2(2)$	2.14–2.11 (m), 1.83–1.81 (m)	25.0	2.14–2.11 (m), 1.83–1.18 (m)	25.0
H–C(3)	4.38 (dd, $J=11.5, 4.5$)	80.8	4.40 (dd, $J=11.5, 4.5$)	81.0
C(4)		42.6		42.6
H–C(5)	1.32 (br. s)	masked	1.33 (br. s)	48.1
$\text{CH}_2(6)$	1.60 (br. s), 1.41–1.38 (m)	18.7	1.61 (br. s), 1.43–1.39 (m)	18.7
$\text{CH}_2(7)$	2.18–2.09 (m), 1.92–1.91 (m)	36.1	2.15–2.12 (m), 1.92–1.91 (m)	35.4
C(8)		42.0		42.2
H–C(9)	1.41–1.38 (m)	51.7	1.43–1.41 (m)	51.6
C(10)		37.8		37.8
$\text{CH}_2(11)$	1.72–1.67 (m), 1.23–1.22 (m)	22.7	1.50–1.47 (m), 1.29 (s)	22.7
$\text{CH}_2(12)$	1.41–1.38 (m), 1.15–1.12 (m)	29.3	1.41–1.38 (m), 1.15–1.12 (m)	28.9
H–C(13)	2.37 (t, $J=7.5$)	33.6	1.42 (br. s)	39.8
C(14)		43.0		43.3
$\text{CH}_2(15)$	1.15–1.11 (m), 1.08–1.05 (m)	28.7	1.15–1.11 (m), 1.02–1.01 (m)	27.8
$\text{CH}_2(16)$	1.41–1.39 (m), 1.28 (br. s)	30.7	1.32–1.28 (m), 1.27 (br. s)	30.5
C(17)		51.9		39.5
H–C(18)	2.08–2.11 (m)	41.3	2.12–2.15 (m)	49.5 (masked)
H–C(19)	1.36 (br. s)	48.1	1.26 (br. s)	32.9
C(20)		148.1		142.4
H–C(21)	5.56 (d, $J=6.5$)	118.3	5.63 (d, $J=6.5$)	122.8
$\text{CH}_2(22)$	1.53 (br. s), 1.34 (br. s)	34.8	1.53 (br. s), 1.34 (br. s)	34.7
$\text{CH}_2(23)$	3.97 (m, ovlp., H_a), 3.77 (d, $J=9.5$, H_b)	70.0	3.99 (d, $J=9.5$, H_a), 3.76 (d, $J=9.0$, H_b)	70.0
Me(24)	0.76 (s)	13.3	0.76 (s)	13.3
Me(25)	0.93 (s)	17.2	0.94 (s)	17.1
Me(26)	0.94 (s)	16.5	1.09 (s)	15.3
Me(27)	1.02 (s)	14.8	1.02 (s)	16.5
H–C(28) or	9.42 (s)	209.0	3.64–3.67 (m, ovlp., H_a), 3.51 (d, $J=11.5$, H_b)	59.2
$\text{CH}_2(28)$				
Me(29)	1.08 (d, $J=6.5$)	23.8	1.01 (d, $J=6.5$)	23.2
$\text{CH}_2(30)$	3.97 (m, ovlp., H_a), 3.94 (d, $J=12.5$, H_b)	64.8	4.21 (ovlp., 2 H)	73.5
Glucose:				
H–C(1')			4.27 (d, $J=8.0$)	104.6
H–C(2')			3.19–3.17 (m)	75.3
H–C(3')			3.33 (masked)	78.1
H–C(4')			3.25 (masked)	71.6
H–C(5')			3.24 (masked)	78.0
$\text{CH}_2(6')$			3.87 (br. d, $J=12.5$), 3.64–3.67 (m, ovlp.)	62.6

Compound **2** was also isolated as a white amorphous powder. The HR-FAB-MS of **2** gave a quasi-molecular-ion peak at m/z 817.3179 ($M - \text{H}]^-$) corresponding to a molecular formula $\text{C}_{36}\text{H}_{58}\text{NaO}_{15}\text{S}_2^-$. The ^{13}C -NMR signals (Table) of **2** were closely related to those of **1**, except for additional signals concerning rings *D* and *E*. Compound **2** differed from **1** by the presence of an additional hexose moiety, and a CH_2OH group

Fig. 1. Important HMBC (H → C) interactions of **1**

instead of a CHO group. The appearance of signals in the $^1\text{H-NMR}$ at $\delta(\text{H})$ 3.64–3.67 (*m*, overlapped, $\text{H}_a\text{-C}(28)$) and 3.51 (*d*, $J = 11.5$ Hz, $\text{H}_b\text{-C}(28)$), and the absence of a CHO signal established the reduction of CHO of **1** to CH_2OH in **2**. This CH_2OH group was unambiguously assigned to the C(28) position. The downfield-shifted signal of $\text{CH}_2(30)\text{OH}$ at $\delta(\text{C})$ 73.5 with the corresponding $^1\text{H-NMR}$ signal at $\delta(\text{H})$ 4.21 (overlapped, 2 H) and the HMBC (Fig. 2) of the anomeric H-atom of the sugar unit at $\delta(\text{H})$ 4.27 (*d*, $J = 8.0$ Hz, $\text{H-C}(1')$) with C(30) ($\delta(\text{C})$ 73.5) showed the attachment of a hexose unit at C(30). The absolute configuration of the sugar unit was assigned as D-glucose after acid hydrolysis of **2** followed by the determination of the optical rotation of the isolated sugar. On the basis of all the above evidences, the structure of **2** was assigned as (3 β ,4 α)-3,23,28-trihydroxyurs-20-en-30-yl β -D-glucopyranoside 3,23-di(sulfate) sodium salt (1:1), which is a new compound.

Fig. 2. Important HMBC (H → C) interactions of **2**

A financial grant from the Higher Education Commission is gratefully acknowledged.

Experimental Part

General. Column chromatography (CC): Polygroprep C-18 (15–25 μm , 10 nm; Macherey–Nagel) and Sephadex LH-20 (25–100 μm ; Sigma–Aldrich). Flash chromatography (FC): Eylea Flash Chromatography EF-10, column (200 mm \times 20 mm i.d.; Eylea) filled with Polygroprep C-18 (25–

40 μm , 10 nm; *Macherey–Nagel*). MPLC: *Eyela-VSP-3050* instrument, column (200 mm \times 25 mm i.d.; *Eyela*) filled with *Polygroprep C-18* (15–25 μm , 10 nm; *Macherey–Nagel*). Optical rotations: *Jasco-DIP-360* automatic digital polarimeter. IR Spectra: *Vector-22* spectrophotometer; $\bar{\nu}$ in cm^{-1} . 1D- and 2D-NMR Spectra: *Bruker-AC-500* and *-AV-600* spectrometers; δ in ppm rel. to Me_4Si as internal standard, J in Hz. FAB-MS: *Jeol-JMS-HX-110* mass spectrometer; in m/z .

Plant Material. The aerial parts of *Z. fabago* were collected from Ankara, Turkey, in June 2002. The plant was identified by one of us (*B. S.*). A voucher specimen (GUE # 2312) was deposited with the Herbarium of the Faculty of Pharmacy, Gazi University, Ankara, Turkey.

Extraction and Isolation. The EtOH extract was prepared from aerial parts (12 kg) of *Z. fabago* by maceration. The dark-green residue (450 g) was dissolved in H_2O and partitioned between hexane, AcOEt, and MeOH. The MeOH extract (4 \times 25 ml) was subjected to CC (*Sephadex LH-20*, pure H_2O (2.0 l), then polarity decrease by MeOH addition in steps of 25% (2.0 l each), up to 100% MeOH (2.0 l)). The fraction (250 mg) obtained with 25% MeOH/ H_2O was subjected to reversed-phase MPLC ($\text{H}_2\text{O}/\text{MeOH}$ 1:1): **1** and **2**. The purity of the compounds was checked by HP-TLC (visualization by spraying with $\text{Ce}(\text{SO}_4)_2$ reagent, followed by heating).

($3\beta,4\beta$)-3,23,30-Trihydroxyurs-20-en-28-al 3,23-Di(sulfate) Sodium Salt (1:1) (**1**): White amorphous powder (12 mg). $[\alpha]_{\text{D}}^{25} = +13.3$ ($c = 0.042$, MeOH). IR (KBr): 3408, 2925, 1710, 1627, 1122. ^1H - and ^{13}C -NMR: *Table*. HR-FAB-MS: 653.2435 ($[M - \text{H}]^-$, $\text{C}_{30}\text{H}_{46}\text{NaO}_{10}\text{S}_2$; calc. 653.2418).

($3\beta,4\alpha$)-3,23,28-Trihydroxyurs-20-en-30-yl β -D-Glucopyranoside 3,23-Di(sulfate) Sodium Salt (1:1) (**2**): White amorphous powder (10 mg). $[\alpha]_{\text{D}}^{25} = +51.4$ ($c = 0.054$, MeOH). IR (KBr): 3418, 2924, 2854, 1625, 1443, 1385, 1200, 1119, 1073, 671, 648. ^1H - and ^{13}C -NMR: *Table*. HR-FAB-MS: 817.3179 ($[M - \text{H}]^-$, $\text{C}_{36}\text{H}_{58}\text{NaO}_{15}\text{S}_2$; calc. 817.3099). FAB-MS: 817 ($[M - \text{H}]^-$), 715, 653, 643.

Acid Hydrolysis. Compound **2** (5 mg) in MeOH (5 ml) was hydrolyzed with 10% aq. HCl soln. for 3 h at 100° . On cooling, the aglycone was extracted with AcOEt. The aq. hydrolyzate was neutralized with Ag_2CO_3 and concentrated. The sugar was found to be glucose by co-TLC (R_f 0.46; AcOEt/MeOH/AcOH/ H_2O 11:2:2:2 and visualization with aniline phthalate reagent). The sugar was identified as D-glucose by the sign of its optical rotation ($[\alpha]_{\text{D}}^{25} = +52.2$).

Detection of Sulfate Group. A 1–2 mg aliquot of compounds **1** and **2** was heated under reflux with 10% HCl soln. (4 ml) for 4 h and then extracted with Et_2O . An aliquot of the aq. layer of each was treated with 70% BaCl_2 soln. to give a white precipitate of BaSO_4 .

REFERENCES

- [1] A. Perrone, M. Masullo, C. Bassarello, A. I. Hamed, M. A. Belisario, C. Pizza, S. Piacente, *J. Nat. Prod.* **2007**, *70*, 584; D. Smati, A.-C. Mitaine-Offer, T. Miyamoto, V. Hammiche, M.-A. Lacaille-Dubois, *Helv. Chim. Acta* **2007**, *90*, 712.
- [2] V. U. Ahmad, Ghazala, S. Uddin, M. S. Ali, *Phytochemistry* **1993**, *33*, 453.
- [3] M. E. O. Matos, M. P. Sousa, M. L. L. Machado, R. B. Filho, *Phytochemistry* **1986**, *25*, 1419.
- [4] T. Mencherini, P. Picerno, C. Scesa, R. Aquino, *J. Nat. Prod.* **2007**, *70*, 1889.
- [5] O. Safir, S. Fkih-Tetouani, N. De Tommasi, R. Aquino, *J. Nat. Prod.* **1998**, *61*, 130.
- [6] V. U. Ahmad, S. S. Khan, A. Ahmed, A. Khan, U. Farooq, S. Arshad, B. Sener, N. Erdemoglu, *Nat. Prod. Commun.* **2007**, *2*, 1085.

Received December 31, 2009